A survey of bacteria found in Belgian dairy farm products

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Description of the subject. Due to the potential hazards caused by pathogenic bacteria, farm dairy production remains a challenge from the point of view of food safety. As part of a public program to support farm diversification and short food supply chains, farm dairy product samples including yogurt, ice cream, raw-milk butter and cheese samples were collected from 318 Walloon farm producers between 2006 and 2014.

Objectives. Investigation of the microbiological quality of the Belgian dairy products using the guidelines provided by the European food safety standards.

Method. The samples were collected within the framework of the self-checking regulation. In accordance with the European Regulation EC 2073/2005, microbiological analyses were performed to detect and count Enterobacteriaceae, Listeria monocytogenes, Salmonella spp., Escherichia coli and Staphylococcus aureus.

Results. Even when results met the microbiological safety standards, hygienic indicator microorganisms like E. coli and S. aureus exceeded the defined limits in 35% and 4% of butter and cheese samples, respectively. Unsatisfactory levels observed for soft cheeses remained higher (10% and 2% for S. aureus and L. monocytogenes respectively) than those observed for pressed cheeses (3% and 1%) and fresh cheeses (3% and 0%) (P ≥ 0.05). Furthermore, the percentages of samples outside legal limits were not significantly higher in the summer months than in winter months for all mentioned bacteria.

Conclusions. This survey showed that most farm dairy products investigated were microbiologically safe. However, high levels of hygiene indicators (e.g., E. coli) in some products, like butter, remind us of applying good hygienic practices at every stage of the dairy production process to ensure consumer safety.

Keywords. Dairy farms, milk products, bacteriological analysis, food safety.

Qualité microbiologique des produits laitiers issus des fermes belges

Description du sujet. Dans le cadre d’un programme public belge de soutien aux fermiers dans la diversification de leurs productions, des échantillons de produits laitiers incluant du yaourt, de la crème glacée, du beurre et du fromage au lait cru ont été prélevés dans 318 fermes en Wallonie, entre les années 2006 et 2014.

Objectifs. Investiger la qualité microbiologique des produits laitiers fabriqués dans les fermes en Belgique.


Résultats. Les résultats obtenus sont conformes aux critères microbiologiques définis. Cependant, dans 35 % des échantillons de beurre et 4 % des échantillons de fromages analysés, les nombres de micro-organismes indicateurs d’hygiène des procédés tels qu’E. coli et S. aureus sont au delà des limites microbiologiques fixées. Le nombre d’échantillons non conformes observé parmi les fromages à pâte molle est plus élevé (10 % et 2 % pour S. aureus et L. monocytogenes) que celui observé parmi les fromages à pâte pressée (3 % et 1 %) ainsi que les fromages frais (3 % et 0 %) (P ≥ 0.05). Par ailleurs, le nombre d’échantillons non conformes est significativement élevé pendant l’été pour toutes les bactéries impliquées.

Conclusions. Cette étude montre que la plupart des produits laitiers étudiés est satisfaisant. Cependant, le nombre élevé de micro-organismes indicateurs d’hygiène (e.g., E. coli) dans certains produits comme le beurre, met en évidence l’importance d’appliquer les bonnes pratiques d’hygiène à toutes les étapes de la production afin d’assurer la sécurité des consommateurs.

Mots-clés. Exploitation laitière, produit laitier, analyse bactériologique, inocuité des produits alimentaires.
1. INTRODUCTION

Milk remains one of the principal products of the dairy sector in Belgium, where the farm cow’s milk production reached approximately 3,474.3 millions kg in 2013 (SPF Économie, PME, Classes moyennes et Énergie, 2014). According to the Belgian Dairy Industry Confederation annual report, the sales of dairy products also increased worldwide in 2012 (CBL, 2013). This implies that hygiene concerns, as well as quality control of dairy foods, will continue and the challenges faced by the dairy industry will increase (Hussein et al., 2005). Cheese is a ready-to-eat food easily contaminated on the surface by undesirable microorganisms. Even if some are spoilage microorganisms, which may produce uncharacteristic visual appearance and diminish the commercial value of the cheeses, others are pathogenic such as Listeria monocytogenes, which have been associated with foodborne listeriosis by consumption of cheese (McLauchlin et al., 2004; Pintado et al., 2010). As the basis of dairy products, raw milk has been shown to be a potential source of food pathogens. In addition, bacteria can enter dairy products at many points during their processing. Such points include entry via the starter culture, floor, packaging material and production room air (Cotton et al., 1992; Kousta et al., 2010; Hill et al., 2012). Numerous foodborne diseases have been associated with milk and dairy products in recent years. According to De Buyser et al. (2001), milk and milk products were implicated in 1–5% of total foodborne disease outbreaks between 1988 and 1998. Langer et al. (2012) reviewed the number of dairy-associated outbreaks during 1993-2006 in the United States and found a total of 121 outbreaks resulting in 202 hospitalizations and 2 deaths. Non-pasteurized as well as pasteurized dairy products associated with these outbreaks included fluid milk and cheeses. Among the causative agents, L. monocytogenes, Salmonella spp., Escherichia coli and Staphylococcus aureus were identified.

Listeria monocytogenes is a ubiquitous pathogen with many possible modes of entry into dairy processing facilities. Parisi et al. (2013) highlighted the wide-spread presence of L. monocytogenes in cheese factories. According to that study, this bacterium can persist for long periods of time, resulting in a continuous contamination of the dairy products. This is of concern because L. monocytogenes has been shown to cause life-threatening disease in foetuses, newborns, immunocompromised people and the elderly (Schuchat et al., 1991).

Salmonella can cause an illness called salmonellosis in humans. In the European Union (EU), over 100,000 cases are reported each year. Moreover an estimated 1 million salmonellosis cases and more than 400 salmonellosis-associated deaths occur annually in the United States (Galans et al., 2006; Scallan et al., 2011; EFSA, 2013; Switt et al., 2013). Salmonella species have been isolated from the faeces of healthy dairy cattle, where they may exist as normal members of the gastrointestinal population (Roy et al., 2001; Wells et al., 2001; Callaway et al., 2005). Moreover, Salmonella has been identified as one of the frequent pathogens associated with foodborne diseases implicated in milk and milk products in France and other countries (De Buyser et al., 2001). Escherichia coli is a bacterium that coexists with its human host in the intestines in a mutually beneficial relationship (Tchaptchet et al., 2011). While most strains of E. coli are commensals, some are known to cause severe enteric disease by infection of the epithelial cells or by the production of toxins. Among them, the E. coli strain O157:H7 is the most frequently involved in human diseases (Caro et al., 2011). Food and dairy products are the main vehicles of E. coli O157:H7, as revealed by a study of 90 outbreaks occurring between 1982 and 2006 in several occidental countries (Snedeker et al., 2009).

The bacterium S. aureus is reported to be one of the most frequent pathogens involved in foodborne diseases associated with dairy products, especially with raw-milk cheese (Techer et al., 2013). The heat-stable enterotoxins produced by this bacterium can cause staphylococcal food poisoning, which ranks as one of the most prevalent causes of gastroenteritis worldwide. Dairy products are frequently involved in food poisoning with enterotoxins levels as low as 0.5 ng·g⁻¹ (Dinges et al., 2000). Moreover, a previous study showed that methicillin-resistant S. aureus was detected in different bovine milks and cheeses marketed in Italy (Normanno et al., 2007; Hennekine et al., 2012). Listeria monocytogenes and Salmonella spp. are included in the European safety criteria Regulation (EC 2073/2005) for dairy products, where defined limits state these microorganisms should be found in “absence in 25 g” or “less than or equal to 100 CFU·g⁻¹” of product. In the same regulation, E. coli and S. aureus are included in the hygienic criteria. Pathogenic E. coli and S. aureus enterotoxins are also included in the safety criteria regulation.

In order to develop or to obtain high added value to its productions and to have a consistent contact with the population, it is important for farm producers to diversify their productions.

In Belgium, the Laboratory of Agro-food Quality and Safety of the University of Liege - Gembloux Agro-Bio Tech, with the support of Public Service of Wallonia (DiversiFerm program), helps and guides the farm producers in their efforts for diversification through hygienic, technological and economic guidance.
According to the European Regulation EC 2073/2005, microbiological analyses should be carried out frequently by the producers to ensure the safety of the food products (self-checking). This is the context in which different data from microbiological analyses have been collected from the farm producers, as part of the self-checking program. The original raw data obtained from the DiversiFerm program include the results from analysis of yogurt, ice cream, raw milk butter and raw milk cheeses. It is important to note that these dairy products were all made in an artisanal way in Walloon farms.

The present work reports a critical evaluation of the data collected over several years. We first investigated the microbiological quality of the dairy products using the guidelines provided by the European food safety standards. We then studied the seasonal distribution of bacteria in the non-complying dairy products from Walloon farms.

2. MATERIALS AND METHODS

Yogurt, ice cream, raw-milk butter and raw-milk cheeses including fresh, soft and pressed cheeses were collected from each of 318 separate farms located throughout the Walloon Region between 2006 and 2014. Fresh cheeses were described as unripened cheeses with short shelf-lives (1-3 weeks), while soft cheeses were aged two months or less. Pressed cheeses, rich in flavor and dry in texture, were the most aged cheeses. Analyses were performed at various stages of their shelf life.

The same sites were sampled at least twice a year, within the self-checking context, and datatotal1,128 dairy product samples were collected. The analyses were conducted by two accredited environmental, toxicology and food control laboratories, in accordance with the European Regulation EC 2073/2005. The detection or enumeration was performed for foodborne pathogens or microorganisms safety indicators like L. monocytogenes and Salmonella spp. and for indicators of hygiene like E. coli and S. aureus. Reference methods were used for the enumeration of Enterobacteriaceae (ISO 21528-2), E. coli (ISO 16649-2) and S. aureus (ISO 6888-2). The S. aureus enterotoxin detection was performed by the method vidas set2 and the kit set-RPLA.

The analyses of L. monocytogenes and Salmonella spp. were performed using validated alternative methods. Analytical methods used for assessing L. monocytogenes levels are either a detection method in 25 g or the colony-count technique in 1 g. When detected, the colony count technique was applied.

All the data were compiled in a Microsoft ACCESS 2007 database and exported to Microsoft Excel 2010 for descriptive and statistical analyses. The minimum, maximum, frequencies and arithmetic mean values were calculated for E. coli, L. monocytogenes and S. aureus. The microbiological quality of dairy products involved in this study was assessed using criteria in the European Regulation EC 2073/2005 (Table 1).

3. RESULTS AND DISCUSSION

3.1. Microbiological quality of cheeses

For this study, 142 pressed cheese, 267 fresh cheese and 92 soft cheese samples were analyzed for S. aureus, E. coli, L. monocytogenes and Salmonella, since these microorganisms present the greatest concern for cheese makers (Johnson et al., 1990; Hill et al., 2013). In 98% of fresh, pressed-type cheeses tested, the microbiological safety and hygiene criteria were achieved, but about 3% and 4% of samples were not within the defined limits regarding to S. aureus and E. coli respectively (Table 2). The cheeses batch where S. aureus levels exceeded the legal limits (10⁵ CFU·g⁻¹), were tested for S. aureus enterotoxins as required by food safety criteria and withdrawn from sale or recalled from the market, even if none of the samples tested contained enterotoxins.

Listeria monocytogenes was present in about 10% of fresh and pressed cheese samples and in 13% of soft cheese samples. The limit of 100 CFU·g⁻¹ was exceeded in 1% pressed cheeses and 2% soft cheeses. Although L. monocytogenes counts exceeding the level of 100 CFU·g⁻¹ were found in relatively few cheese samples (2% and 1% in soft and pressed cheese samples respectively), this microorganism still remains present in all cheese types (Table 3). The significance of L. monocytogenes in cheese and in samples from dairy plants has been previously reported (Makino et al., 2005; Harakeh et al., 2009; Cagri-Mehmetoglu et al., 2011). Even though large prevention methods exist, more continuous monitoring of hygiene measures is needed to avoid spreading in the production facilities when L. monocytogenes is detected.

Staphylococcus aureus criteria levels exceeded limits in 10% of soft cheese samples examined. The counts of this bacterium in about 96% of the cheese samples (all types mingled) analyzed was under the maximum tolerable value M (10⁵ CFU·g⁻¹). The frequent contamination of cheeses or dairy products by S. aureus has been reported by several authors (André et al., 2008; Little et al., 2008; Ostyn et al., 2010; Gücüköüglu et al., 2012). An interesting example of contamination is the one that provoked a food poisoning in Japan in 2000 and that affected 13,420 people due to the ingestion of low-fat milk, contaminated by the enterotoxin (0.08-0.38 ng·ml⁻¹) of S. aureus (Asao et al., 2003). In the present study,
Table 1. Microbiological criteria as recommended by the European Regulation EC 2073/2005 on microbiological criteria for dairy products — Critères microbiologiques applicables aux produits laitiers selon le Règlement européen EC 2073/2005.

<table>
<thead>
<tr>
<th>Product</th>
<th>Microorganisms</th>
<th>Microbiological quality (CFU·g⁻¹)</th>
<th>Satisfactory (&lt; m)</th>
<th>Acceptable (&lt; M)</th>
<th>Unsatisfactory (&gt; M)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Yogurt</td>
<td>Enterobacteriaceae</td>
<td>-</td>
<td>≤ 10</td>
<td>&gt; 10</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Listeria monocytogenes</td>
<td>nd</td>
<td>≤ 10⁶</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
</tr>
<tr>
<td>Ice cream</td>
<td>Enterobacteriaceae</td>
<td>-</td>
<td>≤ 10</td>
<td>&gt; 10</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Listeria monocytogenes</td>
<td>nd</td>
<td>≤ 10⁶</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
</tr>
<tr>
<td>Butter</td>
<td>Escherichia coli</td>
<td>≤ 10</td>
<td>&gt; 10 to 100</td>
<td>&gt; 100</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Listeria monocytogenes</td>
<td>nd</td>
<td>≤ 10⁶</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
</tr>
<tr>
<td></td>
<td>Salmonella spp.</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
</tr>
<tr>
<td>Cheese</td>
<td>Escherichia coli</td>
<td>&lt; 10⁴</td>
<td>10⁴ to &lt; 10⁵</td>
<td>≥ 10⁵</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Staphylococcus aureus</td>
<td>≤ 10⁴</td>
<td>&gt; 10⁴ to &lt; 10⁵</td>
<td>≥ 10⁵</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Listeria monocytogenes</td>
<td>nd</td>
<td>≤ 10⁶</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
</tr>
<tr>
<td></td>
<td>Salmonella spp.</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
</tr>
</tbody>
</table>

m: in a three-class sampling plan, the “m” limit is used to distinguish acceptable quality units (under good manufacturing practices) from those of poor quality — dans un plan d’échantillonnage à trois classes, les unités d’échantillonnage présentant un résultat de moins de « m » sont satisfaits ou de bonne qualité bactériologique; M: represents unacceptable concentrations of microorganisms. Exceeding the “m” level requires corrective action, i.e. revision of the HACCP plan. Exceeding the “M” value requires a recall of the product from the market — les unités révélant un résultat entre « m » et « M » sont jugées comme étant acceptables (médiocres), une révision du plan HACCP s’impose. Et les unités renfermant des comptes supérieurs à « M » sont insatisfaits (non conformes); a: this value is applied to food that does not support growth of L. monocytogenes — la valeur de 100 UFC·g⁻¹ est le seuil maximum admissible de contamination par L. monocytogenes pour la consommation; nd: not detected — non déterminé; -: no data reported — aucune donnée disponible.

Table 2. Overview of microbiological quality of Belgian raw milk cheeses in 114 farms, according to the European Regulation EC 2073/2005 — Qualité microbiologique des fromages au lait cru produits en Belgique dans 114 fermes selon le Règlement européen EC 2073/2005.

<table>
<thead>
<tr>
<th>Cheese type</th>
<th>Total number samples</th>
<th>Bacteria</th>
<th>Microbiological quality (% total samples)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Satisfactory (&lt; m)</td>
</tr>
<tr>
<td>Pressed cheese</td>
<td>142</td>
<td>Escherichia coli</td>
<td>94.4 (134)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Staphylococcus aureus</td>
<td>93.0 (132)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Listeria monocytogenes</td>
<td>88.7 (126)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Salmonella spp.</td>
<td>0.0</td>
</tr>
<tr>
<td>Fresh cheese</td>
<td>267</td>
<td>Escherichia coli</td>
<td>88.8 (237)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Staphylococcus aureus</td>
<td>92.1 (246)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Listeria monocytogenes</td>
<td>93.0 (248)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Salmonella spp.</td>
<td>0.0</td>
</tr>
<tr>
<td>Soft cheese</td>
<td>92</td>
<td>Escherichia coli</td>
<td>84.8 (78)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Staphylococcus aureus</td>
<td>81.5 (75)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Listeria monocytogenes</td>
<td>84.8 (78)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Salmonella spp.</td>
<td>0.0</td>
</tr>
</tbody>
</table>

m, M: see Table 1 — voir tableau 1.
no enterotoxins were found in non-complying samples tested. This is in line with the Kousta et al. (2010) study that found that 96% of the cheese samples (n = 351) collected from different cheese producers conformed to the EU criteria for S. aureus. Furthermore, the levels of unsatisfactory samples observed for fresh cheeses remain lower than those observed for the soft cheeses, considering S. aureus and L. monocytogenes (Figure 1) \((P \geq 0.05)\). Indeed bacterial growth is higher during the cheese ripening process, and it also depends on the specific conditions of cheese varieties such as heat, acid and salt tolerance, initial numbers and individual characteristics of the species or strains in question (Beuvier et al., 2004; Rosengren et al., 2010; Cagri-Mehmetoglu et al., 2011). Lactic fermentation used for fresh cheese production contributes to the reduction of this cheese pH (≤ 4.5), which prevents the development of pathogens. On the other hand, the enzymatic technology contributes to increasing the pH ranges from 4.5 to 4.8 for soft cheeses and from 4.8 to 5.2 for pressed cheeses (24 h after salting). In addition, soft cheese with higher moisture content \((a_w 0.97-0.99)\) provides a more favorable environment for microbial growth than pressed cheese, which presents a moisture content of 0.94-0.97 (Belleflamme et al., 2006; Callon et al., 2011).

Although soft, fresh and pressed cheese samples were examined for the detection of Salmonella spp., none of them were positive for this pathogen. This absence could be explained by maturation conditions, Salmonella characteristics and the adherence to food management regulations (prerequisites and HACCP; Beuvier et al., 2004). Little et al. (2008) also reported no Salmonella in 1,819 cheese samples investigated in UK. This is in contrast with the literature that tells us that a large variety of dairy products, including cheeses, have been linked to human salmonellosis cases (Callaway et al., 2005; Switt et al., 2013).

### 3.2. Microbiological quality of butter

Applying the criteria in the European Regulation EC 2073/2005, 42% of the 363 butter samples examined were satisfactory in their levels of microorganism hygiene indicator \(E. coli\), while 23% of butter samples were acceptable, but 35% exceeded the European Regulation EC 2073/2005 limit for \(E. coli\) (ranging from \(< 1\) to \(10^5\) CFU·g\(^{-1}\); table 4). Several reasons could explain these unsatisfactory samples; for instance, the raw milk used in the butter-making process may not have been subjected to heat treatment, and any viable microorganisms, which may have

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**Table 3.** Distribution of bacterial counts among the positive samples of dairy products analyzed between 2006 and 2014 — Répartition des bactéries au sein des échantillons positifs de produits laitiers analysés entre 2006 et 2014.

<table>
<thead>
<tr>
<th>Dairy product (%)</th>
<th>Bacteria (CFU·g(^{-1}))</th>
<th>Escherichia coli</th>
<th>Staphylococcus aureus</th>
<th>Listeria monocytogenes</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>≤ (10^4)</td>
<td>(10^4) to (10^5)</td>
<td>(≥ 10^5)</td>
<td>≤ (10^4)</td>
</tr>
<tr>
<td>Pressed cheese</td>
<td>94.4</td>
<td>4.2</td>
<td>1.4</td>
<td>93.0</td>
</tr>
<tr>
<td>Fresh cheese</td>
<td>88.8</td>
<td>6.7</td>
<td>4.5</td>
<td>92.1</td>
</tr>
<tr>
<td>Soft cheese</td>
<td>84.8</td>
<td>9.8</td>
<td>5.4</td>
<td>81.5</td>
</tr>
</tbody>
</table>

nd: not detected — non déterminé; \(^{1}\): this value is applied to food that does not support growth of Listeria monocytogenes — la valeur de 100 UFC·g\(^{-1}\) est le seuil maximum admissible de contamination par Listeria monocytogenes pour la consommation.

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**Figure 1.** Schematic representation of unsatisfactory cheeses samples regarding to Escherichia coli, Staphylococcus aureus and Listeria monocytogenes — Représentation schématique des échantillons de fromages contaminés par Escherichia coli, Staphylococcus aureus et Listeria monocytogenes.

The figure shows the percentages of unsatisfactory cheeses samples, analyzed according to the European Regulation EC 2073/2005. The three colors correspond to the percentage of samples which exceed the defined microbiological criteria limits, regarding L. monocytogenes, S. aureus and E. coli — Cette figure montre les pourcentages d’échantillons de fromages non conformes, analysés selon le Règlement européen CE 2073/2005. Les différentes couleurs correspondent au pourcentage d’échantillons contaminés par L. monocytogenes, S. aureus et E. coli au-delà des critères microbiologiques définis.
Bacteria found in Belgian dairy farm products

Table 4. Overview of microbiological quality of Belgian dairy products, according to the European Regulation EC 2073/2005 — Qualité microbiologique des produits laitiers en Belgique, selon le Règlement européen EC 2073/2005.

<table>
<thead>
<tr>
<th>Dairy product</th>
<th>Sample numbers</th>
<th>Farms numbers</th>
<th>Bacteria</th>
<th>Microbiological quality (% total samples)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Satisfactory (&lt; m)</td>
</tr>
<tr>
<td>Yogurt</td>
<td>214</td>
<td>70</td>
<td>Enterobacteria</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Listeria monocytogenes</td>
<td>97.2 (208)</td>
</tr>
<tr>
<td>Ice cream</td>
<td>50</td>
<td>21</td>
<td>Enterobacteria</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Listeria monocytogenes</td>
<td>90.0 (45)</td>
</tr>
<tr>
<td>Butter</td>
<td>362</td>
<td>113</td>
<td>Escherichia coli</td>
<td>42.3 (153)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Listeria monocytogenes</td>
<td>70.4 (255)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Salmonella spp.</td>
<td>0.0</td>
</tr>
</tbody>
</table>

m, M: see Table 1 — voir tableau 1; -: no data reported — aucune donnée reportée.

Originated from a mastitis infection for instance, could have further contaminated the butter.

More frequently, the inadequate cleaning of skimmers could result in a strong growth of *E. coli* during the maturation step (unpublished results). Another source of contamination by *E. coli* could be the butter handlers. To our knowledge, there is less information available on the levels of *E. coli* in raw-milk butter than there is on other dairy products. Considering that pathogenic *E. coli* have been isolated from dairy products (Hussein et al., 2005), the presence of high levels (> 10^4 CFU·g⁻¹) of *E. coli* in some samples tested in our study is a source of concern. Adequate hygienic practices at every stage of the process could help to reduce the health risks linked to these microorganisms, in particular a proper mastitis control program and proper sanitation practices before the maturation of cream starts (milking, skimming and maturation material). *Salmonella* were absent in all butter samples examined and *L. monocytogenes* was present at low levels (ranging from 10 to 40 CFU·g⁻¹) and in only 30% of the samples analyzed. The above results are in accordance with other studies that reported little or no butter samples contaminated by *L. monocytogenes* (Kozak et al., 1996; Aygun et al., 2006). Moreover the *L. monocytogenes* presence in butter is consistent with environmental contaminations, when cleaning and sanitation of material are ineffective.

3.3. Microbiological quality of yogurt

A total of 214 yogurt samples were analyzed for the presence of *L. monocytogenes* and Enterobacteriaceae. As shown in Table 4, all the samples tested were at satisfactory or acceptable levels for *L. monocytogenes*, while 21% were above the legal limits for Enterobacteriaceae. The Enterobacteriaceae counts in unsatisfactory samples ranged from < 10^5 to 10^5 CFU·g⁻¹, and 4 out of 44 samples contained about 10^5–10^6 CFU·g⁻¹. These levels may be due to contamination after pasteurization. According to the European Regulation EC 2073/2005, Enterobacteriaceae are registered as process hygiene indicators in pasteurized milk and other pasteurized dairy products. Therefore, the specific bacteria of this family such as *Salmonella* and *E. coli* were enumerated and researched in the unsatisfactory samples before the yogurt sale.

Normally, yogurt presents a reduced risk of pathogenic bacteria due to the heat treatments applied and the product’s acidic pH of 4.4–4.5 (Morgan et al., 1993; Hill et al., 2013). However, post-pasteurization contamination and the conditions of fermentation such as temperature, fermentation time and final pH are important factors that may affect the presence and survival of unwanted bacteria. Furthermore, the most pathogenic bacterium of the Enterobacteriaceae family, *E. coli* O157:H7, could grow in and survive the acidic conditions of yogurt preparation and thus lead to bacterial enteric infections in consumers (Massa et al., 1997; Gulmez et al., 2003; Cirone et al., 2013). The lack of *L. monocytogenes* in yogurt agrees with Aygun et al. (2006), who found no *L. monocytogenes* in yogurt samples after they examined 157 dairy products. These results confirmed the effort made by the farmers to ensure compliance with the standard, but also and above all an underestimation of the impact of fungi.

3.4. Microbiological quality of ice cream

Among the 50 ice cream samples examined in this study, 30% were above the legal criteria for Enterobacteriaceae (Table 4). The levels of Enterobacteriaceae in ice cream were highly variable, ranging from < 10 to 10^3 CFU·g⁻¹. These Enterobacteriaceae in ice cream could be due
to microorganisms introduced originally by milk, by other ingredients or by the food handlers at the location where these products are prepared. An interesting example of the origin of the ice cream contamination is reported by Fetsch et al. (2014). Indeed, after the food poisoning outbreak caused by staphylococcal enterotoxins in ice cream in Germany, Fetsch and collaborators have identified the equipment used for the production of the ice cream or a contaminated ingredient as the most likely introduction sources of *S. aureus*.

In this study, the ice cream batches where Enterobacteriaceae levels exceed the legal limits (>10) were used for the identification of the various members of Enterobacteriaceae at genus and species level. The non-complying batches were withdrawn from sale. In none of the ice cream samples tested, the levels of *L. monocytogenes* exceeded the 100 CFU·g⁻¹ legal limits, indicating that the microbiological safety criteria were achieved in both cases (Domenech et al., 2013).

4. SEASONAL DISTRIBUTION OF BACTERIA IN DAIRY PRODUCTS

Figure 2 shows the seasonal distribution of unsatisfactory dairy samples analyzed microbiologically from 2006 to 2014. *Listeria monocytogenes*, *E. coli*, *S. aureus* and Enterobacteriaceae have been considered based on their prevalent presence in butter, ice cream, yogurt and cheeses. Even if there are some differences between the percentages of the non-complying dairy products over the seasons, the proportion of these products is not significantly higher (*P* ≥ 0.05) in the summer months than in winter with regards to the mentioned bacteria, by the XLSTAT 2014 test. The proportion of samples contaminated by *L. monocytogenes* beyond the legal limits of 100 CFU·g⁻¹ remained at almost zero across all four seasons. However, statistically significant (*P* < 0.01) seasonal differences in the incidence of *L. monocytogenes* have been found in samples of raw caprine milk (Gaya et al., 1996). According to these authors, the highest incidence occurred in autumn. Several authors have indicated that Shiga-toxin producing *E. coli* excretion by healthy cattle in raw milk varies according to season, peaking in warmer months (Heuvelink et al., 1998; Berry et al., 2010; Farrokh et al., 2013). In the same vein, Caro et al. (2011) have reported that summer is the season with the highest frequency of positive samples for *E. coli* O157.

5. CONCLUSIONS

All analytical results were compared to the safety and hygiene criteria defined by the Regulation EC 2073/2005. All samples analyzed were free of *Salmonella* spp. As regards *L. monocytogenes*, their levels rarely exceeded the legal limit of 100 CFU·g⁻¹. Nevertheless, there is a concern about the presence of *L. monocytogenes* in raw-milk butter, raw-milk cheeses and ice cream samples tested. In order to assess good hygiene practices, hygiene indicators (*E. coli, S. aureus*) were counted in this study. Although the detected levels of bacteria were reasonable in most of
the samples tested, our study highlighted important hygiene issues. The levels of *E. coli* exceeded defined limits in 35% of butter samples and the levels of *S. aureus* exceeded limits in 4% of cheese samples tested. These microorganisms can present a hazard in foods and can affect the health of the consumer. To avoid any health hazards in these artisanal food products, adequate hygienic practices are needed at every stage of the process to assess and control the growth of the bacteria. This survey emphasized the microbiological quality of farm dairy products and provided indications to ensure the safety of the farm food products.

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